



# Advantages of diffuse light for horticultural production and perspectives for further research

Tao Li<sup>1,2\*</sup> and Qichang Yang<sup>1,2\*</sup>

<sup>1</sup> Institute of Environment and Sustainable Development in Agriculture, Chinese Academy of Agriculture Sciences, Beijing, China, <sup>2</sup> Key Laboratory of Energy Conservation and Waste Management of Agricultural Structures, Ministry of Agriculture, Beijing, China

Plants use diffuse light more efficiently than direct light, which is well established due to diffuse light penetrates deeper into the canopy and photosynthetic rate of a single leaf shows a non-linear response to the light flux density. Diffuse light also results in a more even horizontal and temporal light distribution in the canopy, which plays substantial role for crop photosynthesis enhancement as well as production improvement. Here we show some of the recent findings about the effect of diffuse light on light distribution over the canopy and its direct and indirect effects on crop photosynthesis and plant growth, and suggest some perspectives for further research which could strengthen the scientific understanding of diffuse light modulate plant processes and its application in horticultural production.

**OPEN ACCESS** 

#### Edited by:

Edmundo Acevedo, University of California, Davis, USA

### Reviewed by:

Luis Morales-Salinas, University of Chile, Chile Paola Silva, University of Chile, Chile

### \*Correspondence:

Tao Li and Qichang Yang, Institute of Environment and Sustainable Development in Agriculture, Chinese Academy of Agriculture Sciences, Zhongguancun South Street 12, Haidian, 100081 Beijing, China Itao1985@163.com; yangqichang@caas.cn

#### Specialty section:

This article was submitted to Crop Science and Horticulture, a section of the journal Frontiers in Plant Science

Received: 29 June 2015 Accepted: 24 August 2015 Published: 04 September 2015

#### Citation:

Li T and Yang Q (2015) Advantages of diffuse light for horticultural production and perspectives for further research. Front. Plant Sci. 6:704. doi: 10.3389/fpls.2015.00704 Keywords: diffuse light, light distribution, photosynthesis, plant processes, horticultural production

### Introduction

Solar light is composed of a diffuse and a direct component. Diffuse light arises from the scattering of light by molecules or larger particles in the atmosphere and comes from many directions simultaneously; direct light arrives in a straight line from the sun without being scattered (Iqbal, 1983). Plants use diffuse light more efficiently than direct light (Gu et al., 2002; Farquhar and Roderick, 2003; Gu et al., 2003; Alton et al., 2007; Mercado et al., 2009; Li et al., 2014a), it arises due to diffuse light creates a more homogeneous light profile in the canopy than direct light. Photosynthetic rate of a single leaf shows a nonlinear response to the light flux density (Marshall and Biscoe, 1980). High light level usually leads to photosynthetic saturation and decrease in light usually wastes photons by concentrating the light resource to only a fraction of all leaves, leading to a less efficient photosynthetic use of light by plant canopies (Gu et al., 2002). Diffuse light, however, effectively avoids the light saturation constraint by more evenly distributing light among all leaves in plant canopies, and leads to a more efficient use of light (Gu et al., 2002).

To investigate the effect of diffuse light on plant processes, many studies have been carried out by comparing plant responses on cloudy and clear days (Zhang et al., 2011; Urban et al., 2012); or by comparing the aftermath of volcanic and anthropogenic emissions (Gu et al., 2003; Mercado et al., 2009). Such type research implies not only a difference in the fraction of diffuse light, but also large differences in light intensity, and the subsequent changes in microclimatic parameters such as air and soil temperature, and vapour pressure deficit (VPD). These changes directly or indirectly influence plant processes. Recently diffuse glass has become available that increases the diffuseness of light without affecting light transmission in the greenhouse (Hemming et al., 2007, 2008, 2014).

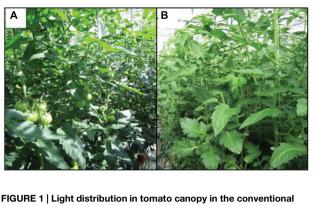
1

Studies have reported that such cover materials have a remarkable effect on plant growth and production (Hemming et al., 2007; Li et al., 2014a,b). Thus, the occurrence of diffuse glass not only provide a promising measure for improving horticultural production, but also offers an opportunity to explicitly explore the pure effects of diffuse light on light distribution over the canopy and its direct and indirect effects on crop photosynthesis and plant growth. In this review, we will discuss the effect of diffuse light on plant processes and its application in horticultural production, and subsequently point out the perspectives for further research.

# Diffusing the Incident Light Improves Spatial Light Distribution

Crop photosynthesis to a large extent correlates with the light profile within the canopy (González-Real et al., 2007; Niinemets, 2007; Sarlikioti et al., 2011a). In the vertical profile of the canopy, light intensity decreases exponentially from top to the bottom of the canopy, as described by the Beer-Lambert-Bouguer law (Chandrasekhar, 1950; Monsi and Saeki, 2005) of which light extinction coefficient can be used to quantify the vertical light distribution in the canopy. Diffuse light exhibits a lower extinction coefficient than direct light (Urban et al., 2012; Li et al., 2014a) although the effect depends on solar position (Morris, 1989). This indicates diffuse light penetrates deeper into the crop canopy. Such phenomenon occurred due to the properties of diffuse light that scatters in many directions and thus causes less shadow, while direct light either concentrates in a beam or casts a shadow in the canopy, which results in the upper leaves being brightly illuminated and lower leaves in deep shade, or strong sunflecks at a given canopy depth. In the horizontal profile of the canopy, diffuse light also results in a more homogeneous light distribution due to less sunflecks occur (Acock et al., 1970; Li et al., 2014a), which plays the most important role for crop photosynthesis enhancement under diffuse light (Li et al., 2014a). A general impression of light distribution in the canopy under direct as well as diffuse light condition has been given in Figure 1. Apart from light distribution, diffuse light also resulted in a lower leaf temperature and less photoinhibition of top leaves (Urban et al., 2012; Li et al., 2014a), which are correlated with the lower light absorption of the top leaves as well as fewer local peaks in light intensity occur under diffuse light, these are also benefit for crop photosynthesis.

Physiological and morphological properties of plant organs can be affected by their prevailing growth microclimate (Sultan, 2000; Niinemets, 2007). A homogeneous light distribution within the crop canopy under diffuse light gives rise to the question whether plant physiological and morphological acclimation occurs. Diffuse light penetrates deeper into the canopy; thus, the lower positioned leaves receive on average a higher light intensity which leads to a higher total nitrogen and chlorophyll content in the canopy, and consequently results in a higher leaf photosynthetic capacity in the lower of the canopy (Li et al., 2014a). Acclimation to diffuse light also includes



clear glasshouse (direct light, A) and diffuse glasshouse (diffuse light, B) on a clear day. Light is more homogeneously distributed under diffuse light (B) compared with direct light (A) where many sunflecks in the middle and lower of the canopy. The photo was taken in Wageningen UR Greenhouse Horticulture, Bleiswijk.

acclimation of leaf morphology, which affects light interception and, consequently, photosynthesis (Pearcy et al., 2005). Li et al. (2014a) reported that tomato plants grown under diffuse light showed a lower specific leaf area (SLA) which indicates a thicker leaves, as well as a higher leaf area index (LAI) which mainly caused by a greater leaf width. A higher LAI is highly relevant for crop photosynthesis, as long as the fraction of light interception is also increased. For the mature crop under greenhouse condition, which often has a closed canopy, thus, the increased LAI under diffuse light has limited effect on canopy light interception and photosynthesis for mature crop (Li et al., 2014a).

# Diffusing the Incident Light Lessens the Variation of Temporal Light Distribution in the Canopy

In nature, temporal light distribution in the canopy is characterized by alternating periods of relatively high light followed by periods of background low light at a given point (sunflecks). Under these circumstances, a large fraction of CO<sub>2</sub> assimilation may occur under transient light conditions. Stomata regulate carbon uptake of a leaf. In response to fluctuating light, stomata exhibit a dynamic response that is slower than the response of photosynthesis and fluctuating light itself, which may limit the CO<sub>2</sub> assimilation under fluctuating light conditions (Pearcy et al., 2004; Lawson and Blatt, 2014). In greenhouses, the shadow and sunflecks generated by overstory leaves, leaf movement, greenhouse construction parts as well as equipment may exacerbate the variation of temporal light distribution. This may substantially limit crop photosynthesis compared to constant light intensities (Pearcy, 1990; Way and Pearcy, 2012). This variation in light intensity can be minimized when the incident light is made diffuse, which would consequently lead to less limitation on leaf photosynthesis, thus improving the canopy LUE (Li et al., 2014b).

Stomatal responses to dynamic light vary dramatically among species, from virtually no response to rapid stomatal responses, thereby resulting in different consequences for instantaneous leaf photosynthesis (Knapp and Smith, 1990; Vico et al., 2011), which may subsequently modulate the effect of diffuse light on canopy LUE. Li (2015) have tested the responses of two anthurium cultivars which have distinct stomatal properties to diffuse light. In cultivars where stomata respond strongly to fluctuations of photosynthetic photon flux density (PPFD), transient rates of photosynthesis and subsequently LUE increased under diffuse light in which stomatal conductance becomes relatively constant and less limiting for photosynthesis. For cultivars with relatively insensitive stomata to the fluctuations of PPFD, the effect of the homogeneous temporal distribution of PPFD on LUE was non-existing. In this context, additional to benefits of diffuse light associated with improved spatial light distribution, the stimulating effect of diffuse light on crop LUE can also depend on the dynamic response of stomatal conductance to incident PPFD at leaf level.

# Allowing More Light via Diffuse Cover Materials Stimulates Growth of Shade-Tolerant Pot Plants without Compromising Plant Quality

Even in northern countries, there are periods in summer with too high light levels for many shade-tolerant pot plants such as anthurium, bromeliads, and orchids. When excessive light energy is being absorbed by the light harvesting antennae at a rate which surpasses the capacity for photochemical and non-photochemical energy dissipation, this may lead to photoinhibition or photodamage (Long and Humphries, 1994). In the long term, this could result in discoloring of leaves or even necrosis. Light damage occurs mostly as a result of prolonged exposure to excessive peaks in light intensity (Asada, 1999; Niyogi, 1999; Kasahara et al., 2002). Consequently, growers regularly apply shading in commercial production of many shade-tolerant pot plants in summer by closing a screen or having white wash on the greenhouse cover. However, shading often carries a penalty on potential crop growth as it is positively related to the amount of light that can be captured, which consequently reduces the LUE in the greenhouse production systems. When diffusing the incident light through cover materials, light in the greenhouse is more homogeneously distributed with less sunflecks, which decreases the extent of photoinhibition as well as local peaks in leaf temperature when global radiation is high (Li et al., 2014a). Therefore, the problem of discoloring of leaves or necrosis in shade-tolerant pot plants under relatively high light could be less when cultivated under diffuse light condition (Li et al., 2014b). Studies have suggested that increasing daily light integral under diffuse light not only accelerates plant growth but also improves plant ornamental quality with more compact plants (Li et al., 2014b; Marcelis et al., 2014). This may substantially contribute to the improvement of horticultural production.

# **Perspectives for Further Research**

Obviously, diffuse light has great advantageous for plant growth. However, detailed studies about the following aspects that closely related with diffuse light are lacking. Further exploring these aspects will strengthen the scientific understanding of diffuse light modulate plant processes as well as its application for crop production.

- (a) The effects of diffuse light on crop photosynthesis could strongly differ between winter and summer light conditions. In winter, photosynthesis of the upper leaves is far from light saturation. With the same light intensity at leaf level, upper leaves have a higher rate of photosynthesis than lower leaves. Therefore, deeper penetration of light may have less effect on crop photosynthesis in winter (Sarlikioti et al., 2011b). Furthermore, light interception follows a seasonal pattern with on average, a lower fraction of light intercepted during summer than during winter because of changes in solar elevation (Sarlikioti et al., 2011a). The higher solar elevation in summer months results in an orientation of light rays more perpendicular to the plant canopy, resulting in a higher light penetration and lower interception. Therefore, seasonal variation of light intensity, directional light quality (diffuse or direct light) as well as solar position should be considered when exploring the effect of diffuse light on light distribution and crop photosynthesis.
- (b) Measuring leaf photosynthesis is the basis for estimating canopy photosynthesis. Conventionally, only the adaxial side of the leaf is illuminated by the light source when measuring single leaf photosynthesis, this might result in minor error in estimating the canopy photosynthesis under diffuse light. This is because diffuseness of light may affect the fraction of light on the abaxial leaf surface, while the abaxial surface have a different photosynthesis light response curve than adaxial surface (Paradiso and Marcelis, 2012). Therefore, measurements of light absorption and photosynthesis light response curves on both the adaxial and abaxial side of leaves in the canopy in combination with functional–structural plant modeling might help to estimate these effects.
- (c) Row crop systems are commonly used in horticultural and agronomic crops. This system facilitates crop management and allows higher light penetration inside the plant canopy. In this system, a fraction of light reaches the ground floor in the middle of the path (Stewart et al., 2003; Sarlikioti et al., 2011a), the reflection of light by the floor can be reused for photosynthesis. Furthermore, row orientation substantially affects canopy light interception (Borger et al., 2010; Sarlikioti et al., 2011a). These effects may differ between diffuse and direct light conditions.
- (d) Light distribution and absorption is highly dependent on crop architecture (Falster and Westoby, 2003; Zheng et al., 2008; Sarlikioti et al., 2011b). Short and compact canopies may generate substantial leaf overlap and self-shading, therefore decreases the net amount of leaf area exposed to light, and consequently affect canopy light interception (Falster and Westoby, 2003). Plants also vary widely in leaf

angle, leaf orientation, internode length, and leaf length to width ratio, these traits have a direct effect on light absorption and photosynthesis (Falster and Westoby, 2003; Sarlikioti et al., 2011b). However, detailed research about plant architecture modulates the effect of diffuse light on light distribution and canopy photosynthesis are lacking. Furthermore, LAI is a predominant factor for canopy light interception, at low LAI mutual shading of leaves within the canopy is small, thus light may readily penetrate deeper into the canopy, which probably decrease the potential effect of diffuse light.

(e) Fruit and vegetable quality is closely correlated with the pre-harvest growth condition. In open field and conventional clear greenhouses, fruit and vegetables often experience diurnal fluctuations or long-term exposure to direct sunlight, with associated high tissue temperatures. This may result in harvest disorders (i.e., sunburn), and heterogeneity of internal quality properties such as sugar content, tissue firmness, mineral content (Woolf and Ferguson, 2000). Fruit with different temperature histories will also respond differently to postharvest low temperatures (i.e., chilling injury) (Ferguson et al., 1999). The quality problems induced by sunlight exposure could be reduced if plants were grown under diffuse light where less fluctuations in temperature and light intensity occurs, detailed research in this aspect has not been reported so far.

### References

- Acock, B. J., Thornley, J. H. M., and Warren Wilson, J. (1970). "Spatial variation of light in the canopy" in Proceedings of the IBP/PP Technical Meeting, Trebon, Czechoslovakia. Wageningen: PUDOC, 91–102.
- Alton, P. B., North, P. R., and Los, S. O. (2007). The impact of diffuse sunlight on canopy light use efficiency, gross photosynthetic product and net ecosystem exchange in three forest biomes. *Glob. Chang Biol.* 13, 776–787. doi: 10.1111/j.1365-2486.2007.01316.x
- Asada, K. (1999). The water-water cycle in chloroplasts: scavenging of active oxygens and dissipation of excess photons. *Annu. Rev. Plant Biol.* 50, 601–639. doi: 10.1146/annurev.arplant.50.1.601
- Borger, C. P. D., Hashem, A., and Pathan, S. (2010). Manipulating crop row orientation to suppress weeds and increase crop yield. *Weed Sci.* 58, 174–178. doi: 10.1614/WS-09-094.1
- Chandrasekhar, S. (1950). Radiative Transfer. London: Oxford University Press.
- Falster, D. S., and Westoby, M. (2003). Leaf size and angle vary widely across species: what consequences for light interception? *New Phytol.* 158, 509–525. doi: 10.1046/j.1469-8137.2003.00765.x
- Farquhar, G. D., and Roderick, M. L. (2003). Pinatubo, diffuse light, and the carbon cycle. Science 299, 1997–1998. doi: 10.1126/science.1080681
- Ferguson, I., Volz, R., and Woolf, A. (1999). Preharvest factors affecting physiological disorders of fruit. *Postharvest Biol. Technol.* 15, 255–262. doi: 10.1016/S0925-5214(98)00089-1
- González-Real, M. M., Baille, A., and Gutiérrez Colomer, R. P. (2007). Leaf photosynthetic properties and radiation profiles in a rose canopy (*Rosa hybrida* L.) with bent shoots. *Sci. Hortic.* 114, 177–187. doi: 10.1016/j.scienta.2007.06.007
- Gu, L., Baldocchi, D. D., Verma, S. B., Black, T. A., Vesala, T., Falge, E. M., et al. (2002). Advantages of diffuse radiation for terrestrial ecosystem productivity. *J. Geophys. Res.* 107, 2–1. doi: 10.1029/2001JD001242
- Gu, L., Baldocchi, D. D., Wofsy, S. C., Munger, J. W., Michalsky, J. J., Urbanski, S. P., et al. (2003). Response of a deciduous forest to the Mount

### Conclusion

Diffuse light improves spatial light distribution in the crop canopy, thereby stimulating crop photosynthesis; the more uniform horizontal light distribution within the canopy plays the most important role for this effect. Diffuse light also lessens the variation of the temporal light distribution at any specific point in the canopy. However, its effect on plant growth depends on the dynamic responses of stomatal conductance to the incident light. Apart from the homogeneous light distribution, diffusing the incident light makes it possible to allow more light in the greenhouse which strongly stimulates crop growth of shade-tolerant pot plants without compromising plant quality. Although the available knowledge have clearly stated the advantageous of diffuse light for crop production, incorporating the seasonal light condition and solar position, plant architecture, crop management practices as well as the post-harvest product quality for further research will strengthen our understanding of the effect of diffuse light on plant processes.

### Acknowledgment

This work was supported by the National High-tech R&D Program of China (863 Program) under contract number 2013AA102407.

Pinatubo eruption: enhanced photosynthesis. Science 299, 2035–2038. doi: 10.1126/science.1078366

- Hemming, S., Dueck, T. A., Janse, J., and van Noort, F. (2007). The effect of diffuse light on crops. Acta Hortic. 801, 1293–1300.
- Hemming, S., Mohammadkhani, V., and Dueck, T. A. (2008). Diffuse greenhouse covering materials-material technology, measurements and evaluation of optical properties. *Acta Hortic*. 797, 469–475.
- Hemming, S., Mohammadkhani, V., and van Ruijven, J. (2014). Material technology of diffuse greenhouse covering materials – influence on light transmission, light scattering and ligh spectrum. *Acta Hortic*. 1037, 883–895.
- Iqbal, M. (1983). An Introduction to Solar Radiation. New York, NY: Academic Press.
- Kasahara, M., Kagawa, T., Oikawa, K., Suetsugu, N., Miyao, M., and Wada, M. (2002). Chloroplast avoidance movement reduces photodamage in plants. *Nature* 420, 829–832. doi: 10.1038/nature01213
- Knapp, A. K., and Smith, W. K. (1990). Contrasting stomatal responses to variable sunlight in two subalpine herbs. Am. J. Bot. 77, 226–231. doi: 10.2307/24 44644
- Lawson, T., and Blatt, M. R. (2014). Stomatal size, speed, and responsiveness impact on photosynthesis and water use efficiency. *Plant Physiol.* 144, 1556–1570. doi: 10.1104/pp.114.237107
- Li, T. (2015). Improving Radiation Use Efficiency in Greenhouse Production Systems. Ph.D. thesis, Wageningnen University, Wageningen.
- Li, T., Heuvelink, E., Dueck, T. A., Janse, J., Gort, G., and Marcelis, L. F. M. (2014a). Enhancement of crop photosynthesis by diffuse light: quantifying the contributing factors. *Ann. Bot.* 114, 145–156. doi: 10.1093/aob/ mcu071
- Li, T., Heuvelink, E., van Noort, F., Kromdijk, J., and Marcelis, L. F. M. (2014b). Responses of two Anthurium cultivars to high daily integrals of diffuse light. Sci. Hortic. 179, 306–313. doi: 10.1016/j.scienta.2014.09.039
- Long, S. P., and Humphries, S. (1994). Photoinhibition of photosynthesis in nature. Annu. Rev. Plant Biol. 45, 633–662. doi: 10.1146/annurev.pp.45.060194.0 03221

- Marcelis, L. F. M., Buwalda, F., Dieleman, J. A., Dueck, T. A., Elings, A., de Gelder, A., et al. (2014). Innovations in crop production: a matter of physiology and technology. *Acta Hortic*. 1037, 39–45.
- Marshall, B., and Biscoe, P. V. (1980). A model for C3 leaves describing the dependence of net photosynthesis on irradiance. J. Exp. Bot. 31, 29–39. doi: 10.1093/jxb/31.1.29
- Mercado, L. M., Bellouin, N., Sitch, S., Boucher, O., Huntingford, C., Wild, M., et al. (2009). Impact of changes in diffuse radiation on the global land carbon sink. *Nature* 458, 1014–1017. doi: 10.1038/nature07949
- Monsi, M., and Saeki, T. (2005). On the factor light in plant communities and its importance for matter production. Ann. Bot. 95, 549–567. doi: 10.1093/aob/mci052
- Morris, J. T. (1989). Modelling light distribution within the canopy of the marsh grass *Spartina alterniflora* as a function of canopy biomass and solar angle. *Agric. For. Meteorol.* 46, 349–361. doi: 10.1016/0168-1923(89)90036-1
- Niinemets, U. (2007). Photosynthesis and resource distribution through plant canopies. *Plant Cell Environ.* 30, 1052–1071. doi: 10.1111/j.1365-3040.2007.01683.x
- Niyogi, K. K. (1999). Photoprotection revisited: genetic and molecular approaches. Annu. Rev. Plant Biol. 50, 333–359. doi: 10.1146/annurev.arplant.50.1.333
- Paradiso, R., and Marcelis, L. E. M. (2012). The effect of irradiating adaxial or abaxial side on photosynthesis of rose leaves. *Acta Hortic*. 956, 157–163.
- Pearcy, R. W. (1990). Sunflecks and photosynthesis in plant canopies. Annu. Rev. Plant Biol. 41, 421–453. doi: 10.1146/annurev.pp.41.060190. 002225
- Pearcy, R. W., Krall, J. P., and Sassenrath-Cole, G. F. (2004). "Photosynthesis in fluctuating light environments," in *Photosynthesis and the Environment*, eds R. Neil and Baker (Amsterdam: Springer), 321–346.
- Pearcy, R. W., Muraoka, H., and Valladares, F. (2005). Crown architecture in sun and shade environments: assessing function and trade-offs with a threedimensional simulation model. *New Phytol.* 166, 791–800. doi: 10.1111/j.1469-8137.2005.01328.x
- Sarlikioti, V., de Visser, P. H. B., and Marcelis, L. F. M. (2011a). Exploring the spatial distribution of light interception and photosynthesis of canopies by means of a functional-structural plant model. *Ann. Bot.* 107, 875–883. doi: 10.1093/aob/mcr006
- Sarlikioti, V., de Visser, P. H. B., Buck-Sorlin, G. H., and Marcelis, L. F. M. (2011b). How plant architecture affects light absorption and photosynthesis in tomato:

towards an ideotype for plant architecture using a functional-structural plant model. Ann. Bot. 108, 1065–1073. doi: 10.1093/aob/mcr221

- Stewart, D. W., Costa, C., Dwyer, L. M., Smith, D. L., Hamilton, R. I., and Ma, B. L. (2003). Canopy structure, light interception, and photosynthesis in maize. *Agron. J.* 95, 1465–1474. doi: 10.2134/agronj2003.1465
- Sultan, S. E. (2000). Phenotypic plasticity for plant development, function and life history. Trends Plant Sci. 5, 537–542. doi: 10.1016/S1360-1385(00)01797-0
- Urban, O., Klem, K., Ač, A., Havránková, K., Holišová, P., Navrátil, M., et al. (2012). Impact of clear and cloudy sky conditions on the vertical distribution of photosynthetic CO2 uptake within a spruce canopy. *Funct. Ecol.* 16, 46–55. doi: 10.1111/j.1365-2435.2011.01934.x
- Vico, G., Manzoni, S., Palmroth, S., and Katul, G. (2011). Effects of stomatal delays on the economics of leaf gas exchange under intermittent light regimes. *New Phytol.* 192, 640–652. doi: 10.1111/j.1469-8137.2011.03847.x
- Way, D. A., and Pearcy, R. W. (2012). Sunflecks in trees and forests: from photosynthetic physiology to global change biology. *Tree Physiol.* 32, 1066– 1081. doi: 10.1093/treephys/tps064
- Woolf, A. B., and Ferguson, I. B. (2000). Postharvest responses to high fruit temperatures in the field. *Postharvest Biol. Technol.* 21, 7–20. doi: 10.1016/S0925-5214(00)00161-7
- Zhang, M., Yu, G. R., Zhuang, J., Gentry, R., Fu, Y. L., Sun, X. M., et al. (2011). Effects of cloudiness change on net ecosystem exchange, light use efficiency, and water use efficiency in typical ecosystems of China. *Agric. For. Meteorol.* 151, 803–816. doi: 10.1016/j.agrformet.2011.01.011
- Zheng, B., Shi, L., Ma, Y., Deng, Q., Li, B., and Guo, Y. (2008). Comparison of architecture among different cultivars of hybrid rice using a spatial light model based on 3-D digitising. *Funct. Plant Biol.* 35, 900–910. doi: 10.1071/FP08060

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2015 Li and Yang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.